

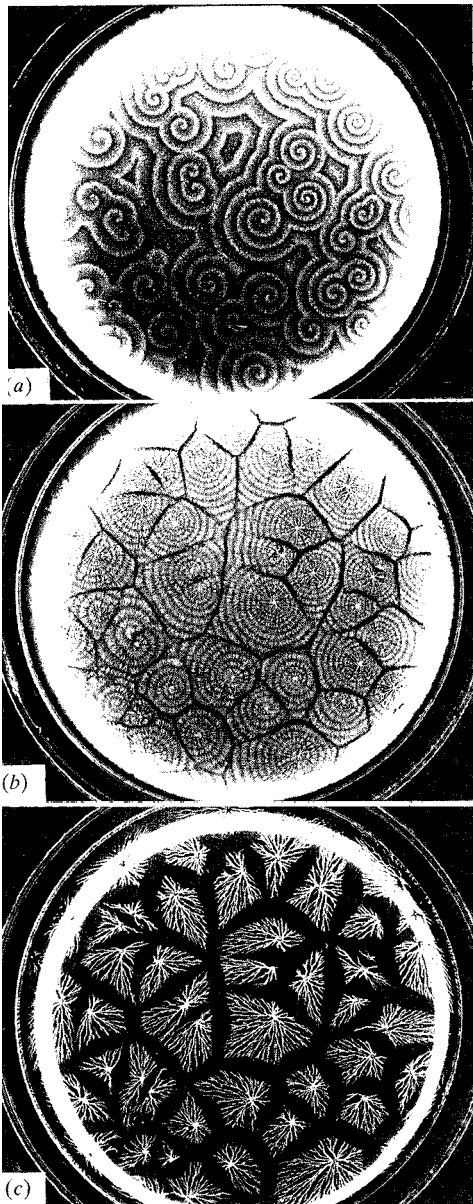


**Self-organization in the
slime mold *Dictyostelium
discoideum***

CS 790R

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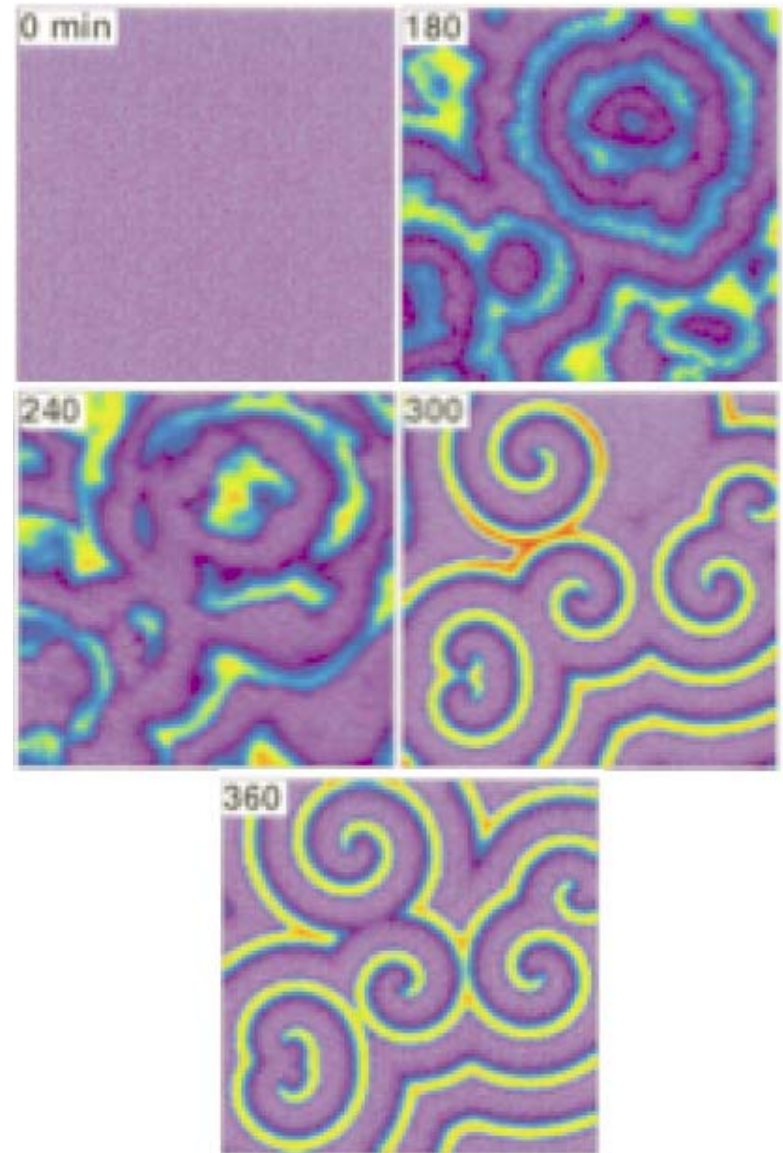
D. Discoideum amoeba starvation response



- Upon starvation: unicellular -> **multicellular (10^4 - 10^5 clumps)**
- **Excitable**: amoebae readily **relay** cyclic Adenosine Monophosphate (cAMP) signals
 - Extracellular cAMP concentrations trigger the production and release of cAMP as **positive feedback**
 - Wave and spiral patterns of cAMP form
- **Chemotaxis**: amoebae travel up cAMP gradients; (a) spiral pattern emerges (b,c) and amoebae **stream** towards spiral centers
- Eventually form “slug,” differentiate (e.g. we notice cell types), produce spores
- But is it self-organization?

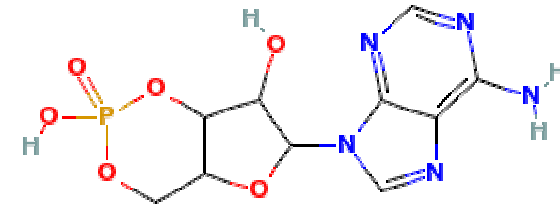
The evidence for self-organization

- Each amoeba must be responding to its **local environment**
- A **global pattern** formed
- Wave “speed & period not generated by a master amoeba,” according to experiments
- Could there be *de facto* pacemakers?
 - Those that starve first, by chance



cAMP-secretion: Oscillation and Relay

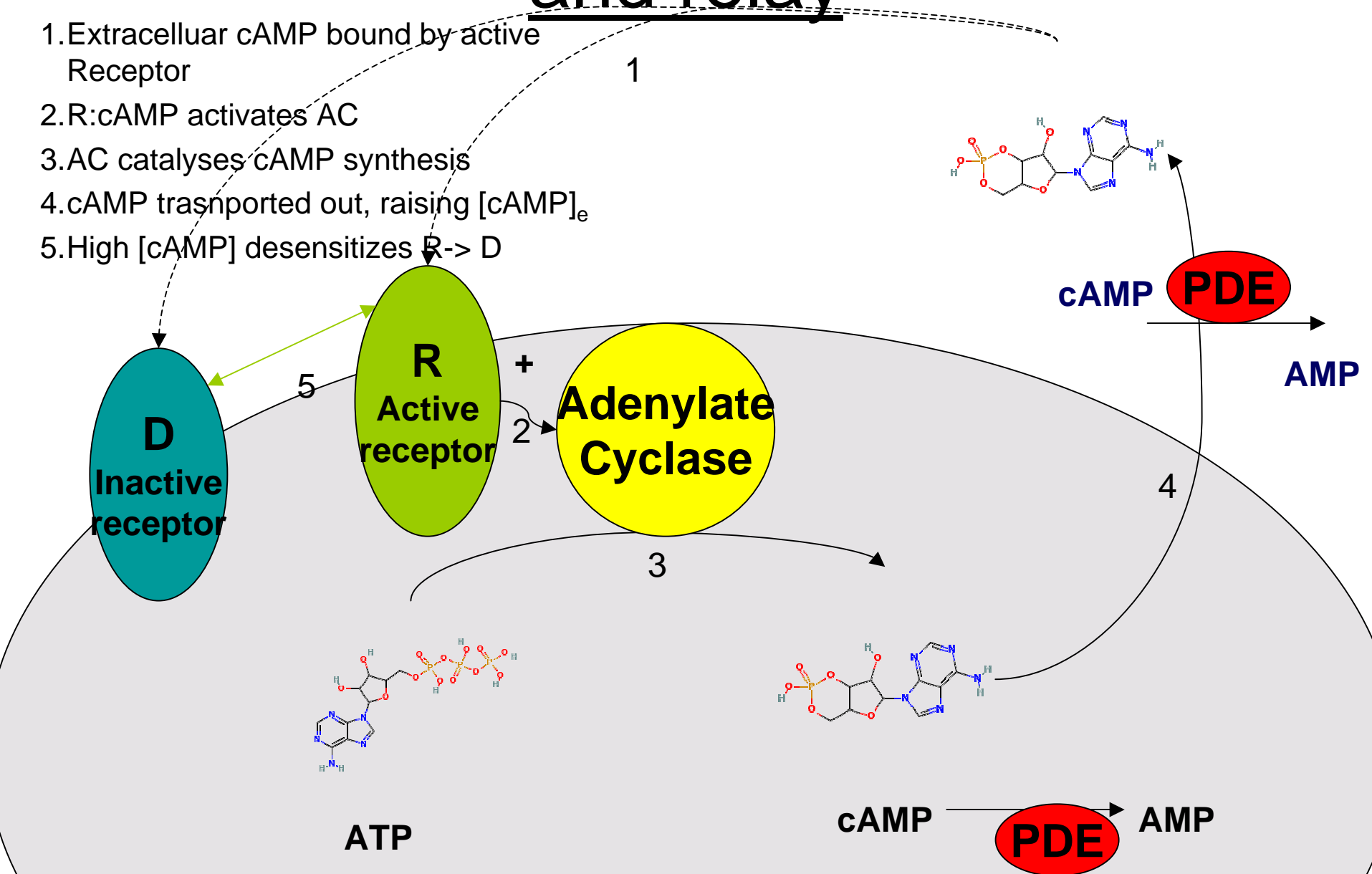
- Oscillatory release
 - Stirred & starved cell suspensions release cAMP in **5-10 minute pulses**
- Relay
 - If a large enough cAMP pulse added to medium...
 - Cells amplify pulse (**positive feedback**)



Cyclic AMP

Basic schematic of cAMP oscillation and relay

1. Extracellular cAMP bound by active Receptor
2. R:cAMP activates AC
3. AC catalyses cAMP synthesis
4. cAMP transported out, raising $[cAMP]_e$
5. High $[cAMP]$ desensitizes R \rightarrow D



Modeling cAMP waves

Only three variables needed to model secretion

$$\frac{\partial \gamma}{\partial t} = (k_t \beta / h) - k_e \gamma + D_\gamma \nabla^2 \gamma \quad = \text{rate of change of } \underline{\text{extracellular [cAMP]}}$$

Secretion	-degradation	+ diffusion
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$$\frac{\partial \beta}{\partial t} = \phi(\rho, \gamma) - k_i \beta - k_j \beta \quad = \text{rate of change of } \underline{\text{intracellular [cAMP]}}$$

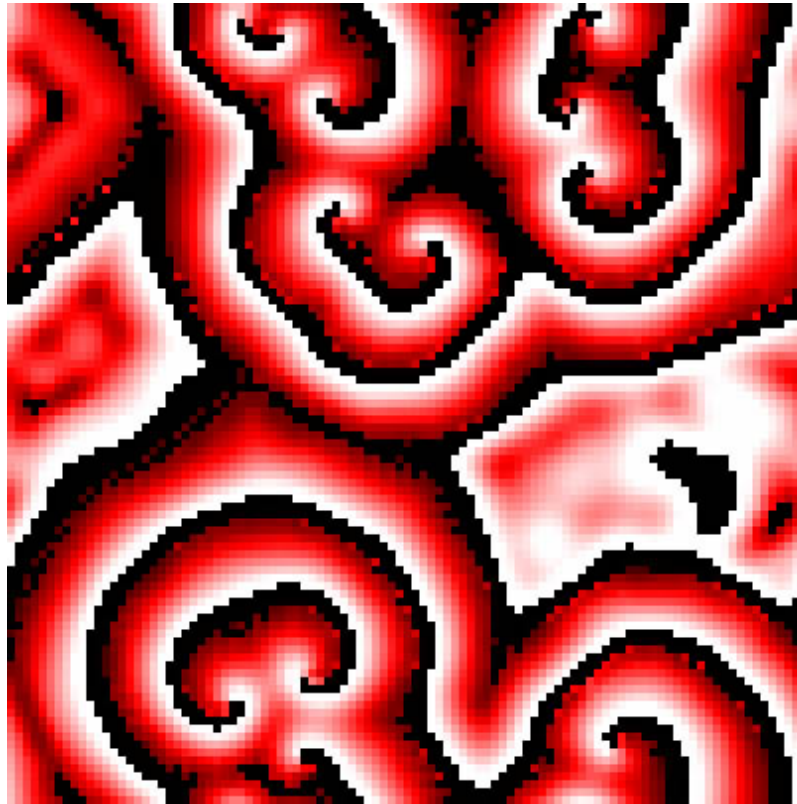
Synthesis	-secretion	-degradation
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$$\frac{d\rho_T}{dt} = -f_1(\gamma)\rho_T + f_2(\gamma)(1 - \rho_T) \quad = \text{rate of change of } \underline{\text{active receptors}}$$

-desensitization	+ resensitization
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NetLogo simulation: BZ

BZ-reactions provide a rough analogy of the cAMP as excitable waves



Modeling cell movement:

- New assumptions
 - Ea. cell detects cAMP gradient & moves up it
 - Prolonged cAMP stimulus desensitizes ability to detect and/or respond to gradients
 - Figure 8.10
 - receptor-mediated desensitizations as before?
 - Polarized motor machinery?
 - Cell-cell adhesions stymie clump dispersal
- $\frac{\partial \gamma}{\partial t}$ is now also a function of local cell density
 - (p.111 if you're interested)

Modeling cell movement: the math

- *Cell diffusion*

- Variable diffusion coefficient

$$\mu(n) = \mu_1 + \mu_2 N^4 / (N^4 + n^4)$$

- Cell density & threshold

- *Chemotaxis*

- Proportional to [cAMP], density (n), and desensitization (p. 115):

$$\chi(\rho) = \frac{\chi_0 \rho^m}{A^m + \rho^m}$$

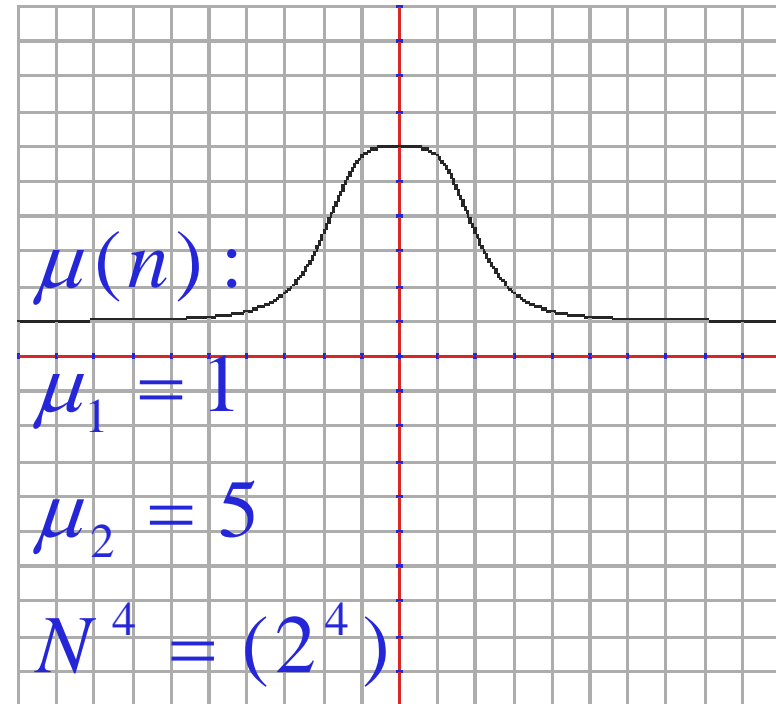
ρ = active Receptor proportion

A, m = constants

- “They” emphasize shape

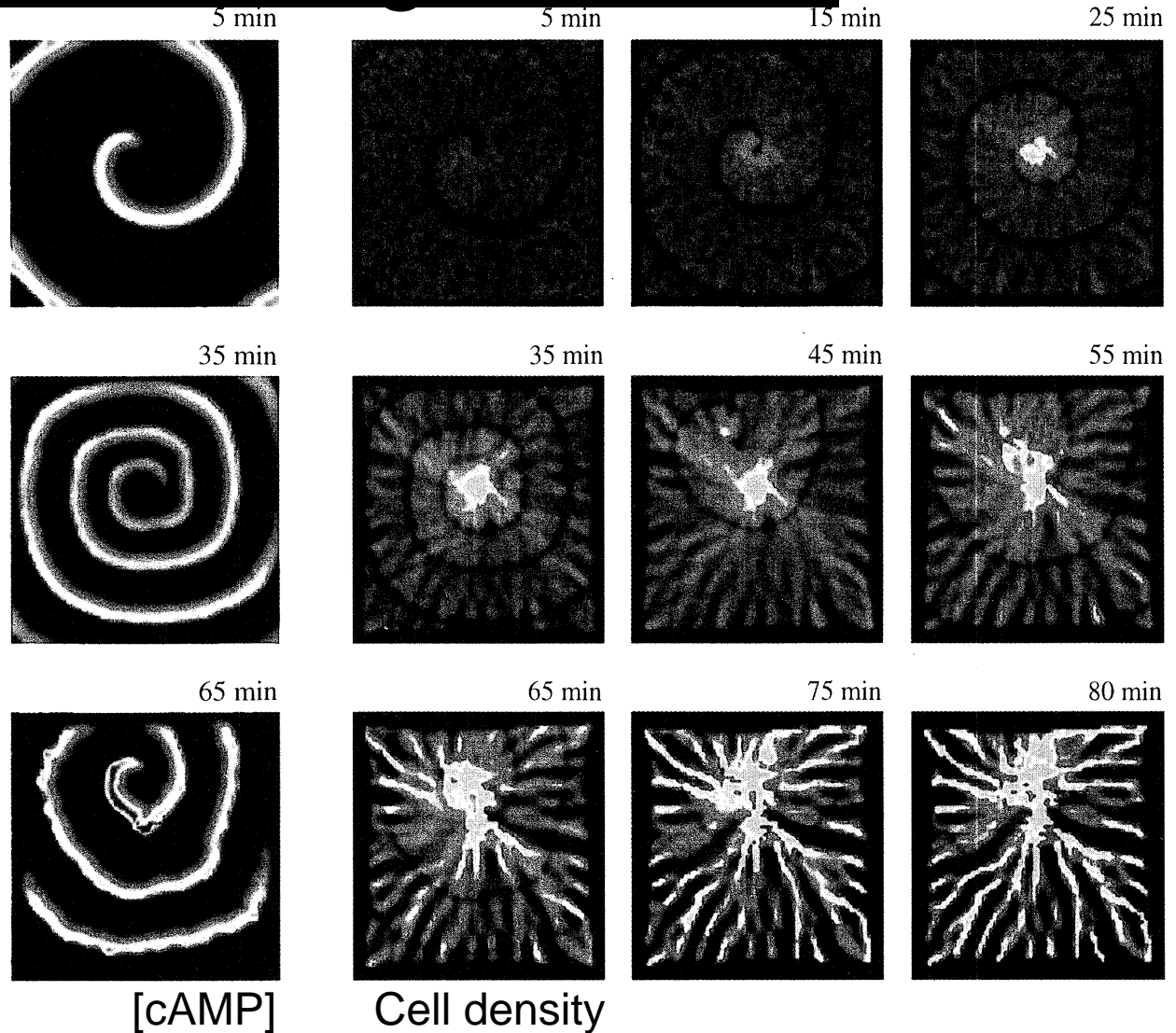
over specifics here

$$\frac{\partial n}{\partial t} = \nabla \cdot [\mu(n) \nabla n] - \nabla \cdot [\chi(\rho) n \nabla u]$$



As $n \rightarrow N$, $\mu(n)$ falls sharply

Correlating cAMP patterns with streaming behavior



Streaming slime “simulation”

